

AccuPOL™ DNA Polymerase

With 10x Ammonium Buffer (15 mM MgCl₂)

Concentration: 2.5 units/µl

Cat. No.: A211104 1000 Units

ooo omts			
-	AccuPOL DNA Polymerase	10x Ammonium Buffer, 15 mM MgCl ₂	MgCl₂ 25 mM
ID No.	5101800	5100950	5575801
Cap colour	Blue	White	Clear
Content	2 x 200 μl	2 x 1.5 ml	2 x 1.5 ml

Key Features

AccuPOL DNA Polymerase is a thermostable enzyme with proof-reading ability, which can be used in primer extension reactions and other molecular biology applications. AccuPOL exhibits $5'\rightarrow 3'$ DNA polymerase activity and $3'\rightarrow 5'$ proofreading exonuclease activity. The latter allows the enzyme to correct misincorporated nucleotides. AccuPOL has an error rate* of 1.1×10^{-6} , which gives a $16 \times 16 \times 16^{-6}$ greater fidelity than Taq Polymerase. Optimal reaction conditions are achieved by using the 10×16^{-6} Ammonium buffer containing MgCl₂. AccuPOL DNA Polymerase is recommended for applications, which require high fidelity or blunt ending.

* The error rate is measured using the LacIOZ assay. Fidelity depends also on reaction conditions.

Kit Components

AccuPOL DNA Polymerase in Storage Buffer

2.5 u/ μ l AccuPOL DNA Polymerase, 50 mM Tris-HCl pH 8.5, 0.1 mM EDTA, 1.0 mM DTT, 0.1% Tween $^{\otimes}$ 20, 50% Glycerol.

10x Ammonium Buffer

Tris-HCl pH 8.5, $(NH_4)_2SO_4$, 15 mM MgCl₂, 1% Tween[®] 20.

Ammonium in the buffer minimizes the need for optimization of the MgCl_2 concentration or the annealing temperature for most primer-template systems.

MgCl₂

25 mM MgCl₂ in PCR grade water.

Recommended Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label.

Option: Store at +4 °C for up to 6 months.

Quality Control

AccuPOL DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity or nicking activity.

Unit Definition

One unit is defined as the amount of enzyme that incorporates 10 nmol of dNTPs into an acid-precipitable form of DNA in 30 minutes at 72 $^{\circ}\text{C}$ under standard assay conditions.

Protocol

This protocol serves as a guideline to ensure optimal PCR results when using AccuPOL DNA Polymerase. Optimal reaction conditions such as incubation times, temperatures and amount of template DNA may vary and must be determined individually.

- Thaw 10x Buffer, dNTP mix, and primer solutions. It is important to thaw the solutions completely (some buffers need to reach room temperature) and mix thoroughly before use to avoid localized concentrations of salts. Keep all components on ice. The polymerase is provided in glycerol and does not need thawing. Keep it at -20 °C at all times.
- Prepare a master mix according to Table 1. The master mix typically contains all the components needed for extension except the template DNA. We recommend Ampliqon Ammonium Buffer to be used with AccuPOL Polymerase.

Important: It is critical to withhold AccuPOL Polymerase until after addition of dNTPs. Otherwise, the proofreading activity of the polymerase may degrade the primers resulting in non-specific amplification and reduced product yield.

Table 1. Reaction components (master mix and template DNA)

Component	Vol./reaction*	Final concentration*
10x Buffer	5 μΙ	1x
25 mM MgCl ₂	0 μl (0 – 6 μl)	1.5 mM (1.5 – 4.5 mM)
dNTP mix (12.5 mM each)	0.8 μΙ	0.2 mM of each dNTP
Primer A (10 μM)	1 μΙ (0.5 – 5 μΙ)	0.2 μΜ (0.1 – 1.0 μΜ)
Primer B (10 μM)	1 μΙ (0.5 – 5 μΙ)	0.2 μΜ (0.1 – 1.0 μΜ)
AccuPol DNA Pol.	0.6 μl (0.6 – 2 μl)	1.5 units (1,5 – 5 units)
PCR-grade H ₂ O	Χ μΙ	-
Template DNA	ХμΙ	genomic DNA: 50 ng (10 – 500 ng) plasmid DNA: 0.5 ng (0.1 – 1 ng) bacterial DNA: 5 ng (1 – 10 ng)
TOTAL volume	50 μΙ	-

^{*} Suggested starting conditions; theoretically used conditions in brackets. The final volume can be reduced to 25 μ l by using half of the volumes suggested in Vol./reaction, eg. 0.3 μ l AccuPol instead of 0.6 μ l AccuPol.

- 3. Mix the master mix thoroughly and dispense appropriate volumes into reaction tubes. Mix gently, e.g. by pipetting the master mix up and down a few times.
- Add template DNA to the individual tubes containing the master mix.
- Program the thermal cycler according to the manufacturer's instructions. AccuPOL is a proofreading enzyme and require an extension time of 1 – 2 min/kb.

For maximum yield and specificity, temperatures and cycling times should be optimized for each new template target or primer pair.

6. Place the tubes in the thermal cycler and start the reaction.

MADE IN **DENMARK**

Three-step PCR program

Cycles	Duration of cycle	Temperature
1	1 – 2 minutes ^a	95 ℃
25 - 35	30 – 60 seconds ^b	95 ℃
	30 seconds ^c	50 − 65 °C
	1 – 4 minutes ^d	72 °C
1	5 minutes ^e	72 °C

a. Initial denaturation step.

- ^c Annealing step: The reaction temperature is lowered to 50-65 °C for 30 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3-5 °C below the T_m (melting temperature) of the primers used.
- d. Extension/elongation step: The extension rate of AccuPOL DNA polymerase is slower than that of Taq DNA Polymerase. Therefore, during the extension step, allow approximately 2 minutes for every 1kb to be amplified (minimum extension time of 1 minute).
- ^{a.} Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

Notes:

■ 15 mM MgCl₂ is present in 10x PCR Buffer. The 1x concentration is 1.5 mM MgCl₂. In some applications, more than 1.5 mM MgCl₂ is required for best results. For this reason, 25 mM MgCl₂ is included in the kit. Table 2 provides the volume of 25 mM MgCl₂ to be added to the master mix if a higher MgCl₂ concentration is required.

Table 2. Additional volume (μl) of MgCl₂ per 50 μl reaction

Final MgCl ₂ conc. in reaction (mM)	1.5	2.0	2.5	3.0	3.5	4.0	4.5
Volume of 25 mM MgCl ₂	0	1	2	3	4	5	6

 For longer DNA targets more DNA polymerase could be added to the PCR master mix.

Related Products

AccuPOL™ DNA Polymerase (500 units)	Cat. No.
AccuPOL™ DNA Polymerase 2.5 U/μl	A210003
• with 10x Ammonium Buffer	A211103

AQ90 High Fidelity DNA Polymerase (500 units)	Cat. No.
AQ90 High Fidelity DNA Polymerase 2 U/μl • with 10x Ammonium Buffer	A457403
AQ90 High Fidelity Master Mix (500 x 50 μl)	Cat. No.
2x Master Mix, 2 mM MgCl ₂ final concentration	A470703

Taq DNA Polymerase (500 units) *	Cat. No.
Taq DNA Polymerase 5 U/μl	A110003
with 10x Ammonium Buffer	A111103
Hot Start DNA Polymerase (500 units) *	Cat. No.
TEMPase Hot Start DNA Polymerase, 5 U/μl	A220003
with 10x Ammonium Buffer	A221103

^{*}Available in kits including one or two buffers (Ammonium Buffer, Standard Buffer or Combination Buffer). All kits include extra $25\,\text{mM}$ MgCl $_2$

Ultrapure dNTPs*	Cat. No.
dNTP Mix 40 mM (2 x 500 μl): 10 mM each dA, dC, dG, dT	A502004
dNTP Set, 100 mM each: 250 μl of each dA, dC, dG and dT	A511104

^{*}Other concentrations and Single dNTPs are available.

Loading Buffers, PCR water and Ladders	Cat. No.
5x Loading Buffer Red *, 5 x 1 ml	A608104
Iqon PCR Ladder **, 100 – 3000 bp, 1 x 0.5 ml	A610341
PCR Grade Water, 6 x 5 ml	A360056

^{*} Also available with Blue, Orange or Cyan. ** Available in different size ranges.

Buffer for AccuPol polymerase *	Cat. No.
10x Ammonium Buffer, 3 x 1.5 ml	A301103
Betaine Enhancer Solution 5 M, 5 x 1 ml	A351104

^{*}Ammonium Buffer is also available as ${\rm Mg}^{2^+}$ free buffer, detergent free buffer and ${\rm Mg}^{2^+}$ and detergent free buffer. **For direct gel loading and visualisation.

For Research Use Only. Not for use in diagnostics procedures.

Other product sizes, combinations and customized solutions are available. Please look at www.ampliqon.com or ask for our complete product list for PCR Enzymes. For customized solutions please contact us.

Made in Denmark Issued 08/2021

b. Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 30 – 60 seconds. It causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.